RA213-EN.01

Human Papillomavirus Type 16 (HPV16) Specific ELISA Kit

Pack Size: 96 tests

Catalog Number：RAS-A213

IMPORTANT: Please carefully read this manual before performing your experiment.

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedures

INTENDED USE

The kit is developed for quantitative detection of HPV16 L1 in samples. It is intended for research use only (RUO).

PRINCIPLE OF THE ASSAY

Human papillomavirus (HPV) is the major cause of cervical cancer and genital warts, which occur through the persistent infection of the proliferating cells of the epithelium. HPV is a non-enveloped, double-stranded DNA virus with a T = 7 icosahedral capsid composed of 72 L1 pentamers (capsomeres) paired with L2 monomers. HPV L1-only virus-like particles (VLPs) have been obtained from different expression systems and can induce a strong immune response against HPV infection. Currently, commercially available prophylactic HPV vaccines are in the form of VLP.

A rapid and effective assay kit detecting the levels of HPV L1 is urgently needed to accelerate the development of HPV vaccines.

This assay kit is used to measure the levels of HPV16 L1 by employing a standard sandwich-ELISA format. The microplate in the kit has been pre-coated with Anti-HPV16-L1 Antibody. First add the standard samples provided in the kit and your samples to the plate, incubate and wash the wells. Then add the HRP-Anti-HPV16-L1 Antibody to the plate, incubate and wash the wells. Lastly load the substrate into the wells and monitor color development in proportion with the amount of HPV16 L1 present. The reaction is stopped by the addition of a stop solution and the intensity of the absorbance can be measured at 450 nm and 630 nm. The OD Value reflects the amount of HPV16 L1 bound.

MATERIALS PROVIDED

TABLE 1. MATERIALS PROVIDED

Catalog	Components	Size (96 tests)	Format	Storage	
				Unopened	Opened
RAS213-C01	Pre-coated Anti-HPV16-L1 Antibody Microplate	1 plate	Solid	2-8℃	2-8℃
RAS213-C02	HPV16-L1 Standard	15 μg	Powder	2-8℃	-70℃
RAS213-C03	HRP-Anti-HPV16-L1 Antibody	15 μg	Powder	2-8℃, avoid light	-70℃, avoid light
RAS213-C04	10×Washing Buffer	50 mL	Liquid	2-8℃	2-8℃

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RAS213-C05	Sample Dilution Buffer	50 mL	Liquid	2-8℃	2-8℃
RAS213-C06	HRP-Antibody Dilution Buffer	25 mL	Liquid	2-8℃	2-8℃
RAS213-C07	Substrate Solution	12 mL	Liquid	2-8℃, avoid light	2-8℃, avoid light
RAS213-C08	Stop Solution	7 mL	Liquid	2-8℃	2-8℃

Note: *The expression system of HPV16-L1 is Hansenula polymorpha.*

REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED

Single or dual wavelength microplate reader with 450 nm and 630 nm filter;

Centrifuge;

37ºC Incubator;

10 µL, 200 µL and 1000 µL precision pipettes;

10 µL, 200 µL and 1000 µL pipette tips;

Multichannel pipettes;

Tubes;

Graduated cylinder to prepare Wash Solution;

Deionized or distilled water to dilute 10×Washing Buffer;

STORAGE

1. Unopened kit should be stored at 2℃-8℃ upon receiving.

2. Find the expiration date on the outside packaging and do not use reagents past their expiration date.

3. The opened kit should be stored per components table. The shelf life is 30 days from the date of opening.

REAGENT PREPARATION

1. Bring all reagents and samples to room temperature (20℃-25℃) before use. If crystals have formed in buffer solution, place the sample in a 37 ºC incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.

2. Reconstitute the provided lyophilized materials to stock solutions with distilled, sterile water as recommended in Table 2 and place the materials for 15 to 30 minutes at room temperature with occasional gentle mixing. Avoid

vigorous shaking. The reconstituted stock solutions should be stored at -70°C. It is recommended not to freeze-thaw more than 1 time, the packing specification shall not be less than 4 µg.

TABLE 2. RECONSTITUTION METHODS FOR 96 TESTS

ID	Components	Size	Stock Solution Con.	Reconstitution Buffer and Vol.
RAS213-C02	HPV16-L1 Standard	15 µg	100 µg/mL	150 µL water
RAS213-C03	HRP-Anti-HPV16-L1 Antibody	15 µg	100 µg/mL	150 µL water

RECOMMENDED SAMPLE PREPARATION

1. Working fluid preparation

1.1 Preparation of 1×Washing Buffer:

Dilute 50 mL 10×Washing Buffer with ultrapure water/deionized water to 500 mL.

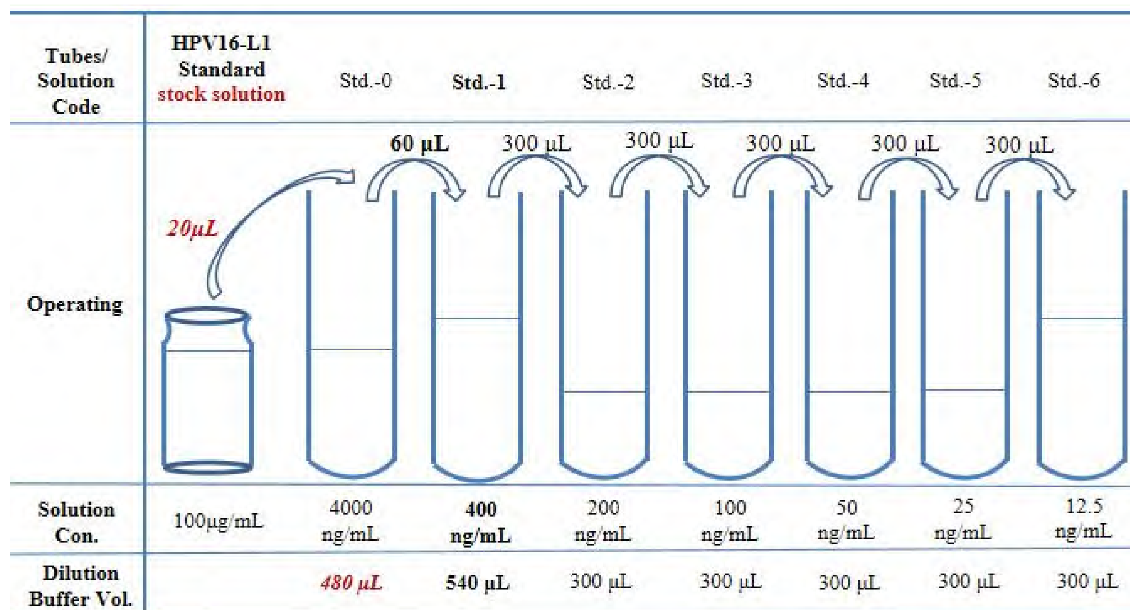
1.2 Preparation of HRP-Anti-HPV16-L1 Antibody working fluid:

Dilute HRP-Anti-HPV16-L1 Antibody to 0.8 µg/mL with HRP-Antibody Dilution Buffer. The prepared working fluid should avoid light. Please prepare it for one-time use only.

2. Preparation of Standard Curve

Make serial dilutions of the HPV16-L1 as a Standard curve with Sample Dilution Buffer as recommended in Figure 1.

FIGURE 1. PREPARATION OF 1:1 SERIAL DILUTIONS OF THE HPV16-L1



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3. Add Samples

Add 100μL serially diluted **HPV16-L1 Standard** curve and samples to each well. For blank Control wells, please add 100μL Sample Dilution Buffer. Seal the plate with microplate sealing film and incubate at 37℃ for 1.0 hour.

Note: *It is recommended to set double holes for samples and standard curves to be tested.*

4. Washing

Remove the remaining solution by aspiration, add 300 μL of 1×Washing Buffer to each well, soak for 30s, remove any remaining 1×Washing Buffer: by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

5. Add HRP-Anti-HPV16-L1 Antibody

For all wells, add 100 μL **HRP-Anti-HPV16-L1 Antibody (dilute to 0.8 μg/mL)** working solution. Seal the plate with microplate sealing film and incubate at 37℃ for 1.0 hour.

6. Washing

Repeat step 4.

7. Substrate Reaction

Add 100 μL **Substrate Solution** to each well. Seal the plate with microplate sealing film and incubate at 37℃ for 20 min, avoid light.

8. Termination

Add 50 μL **Stop Solution** to each well and tap the plate gently to allow thorough mixing.

Note: *the color in the wells should change from blue to yellow.*

9. Data Recording

Read the absorbance at 450 nm and 630 nm using UV/Vis microplate spectrophotometer within 5 minutes.

Note*: To reduce the background noise, subtract the value read at* $OD_{450\ nm}$ *with the value read at* $OD_{630\ nm}$.

CALCULATION OF RESULTS

1. Normal range of Standard curve: $R^2 \geq 0.9900$, detection range: 12.5-400 ng/mL.

2. If the OD value of the sample to be tested is higher than the highest standard, the sample shall be diluted with dilution buffer and assay repeated.

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3. To calibrate absorbance value obtained by the standard curve, the OD value of the sample to be measured is subtracted from the OD value of the blank control. The standard curve is plotted with the standard concentration as x-axis and the calibrated absorbance value as y-axis. Four parameters logistic are used to draw the standard curve and calculate the sample concentration.

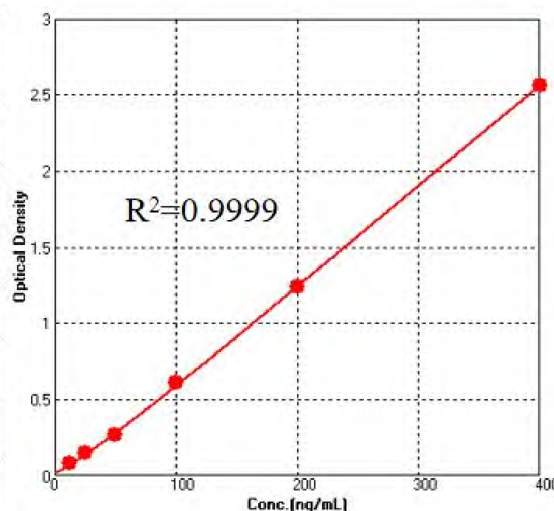
PRECAUTIONS

1. This kit is for research use only and is not for use in diagnostic or therapeutic procedures.
2. The kit should be used according to the instructions.
3. Do not mix reagents from different lots.
4. Bring all reagents and samples to room temperature (20℃-25℃) before use. If crystals have formed in buffer solution, warm to room temperature until the crystals have completely dissolved.
5. The kit should be stored at 2°C to 8°C.

TYPICAL DATA

The following data is for reference only. The sample concentration was calculated based on the results of the standard curve.

Standard (ng/mL)	O.D.-1	O.D.-2	Average	Corrected
400	2.653	2.576	2.615	2.565
200	1.324	1.258	1.291	1.242
100	0.649	0.663	0.656	0.607
50	0.310	0.323	0.317	0.267
25	0.202	0.190	0.196	0.147
12.5	0.136	0.117	0.127	0.077
0	0.049	0.050	0.050	/



PRECISION

1. Intra-assay Precision

Three samples of known concentration were tested ten times on one plate to assess intra-assay precision.

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2. Inter-assay Precision

Three samples of known concentration were tested in three separate assays to assess inter-assay precision.

	Intra-assay Precision			Inter-assay Precision		
Sample	1	2	3	1	2	3
n	10	10	10	3	3	3
Mean (ng/mL)	265.549	131.489	23.094	263.789	132.975	24.273
SD	14.112	5.226	1.580	1.762	3.470	1.513
CV (%)	5.3	4.0	6.8	0.7	2.6	6.2

***Note**: The example data is for reference only.*

RECOVERY

Three samples with different concentrations were tested to calculate the recovery rate.

Sample(n=5)	Average Recovery %	Range %
High	90.0	82.5-100.9
Middle	86.1	80.1-92.3
Low	96.1	82.0-118.9

LINEARITY

To assess the linearity of the assay, samples spiked with high concentrations were serially diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

		Cell culture medium (DMEM)	Cell culture medium (1640)
1:2	Average Recovery (%)	92.9	102.9
	Range (%)	81.4-112.7	95.3-109.3
1:4	Average Recovery (%)	89.8	98.3
	Range (%)	82.0-98.5	87.5-108.0
1:8	Average Recovery (%)	91.0	93.0

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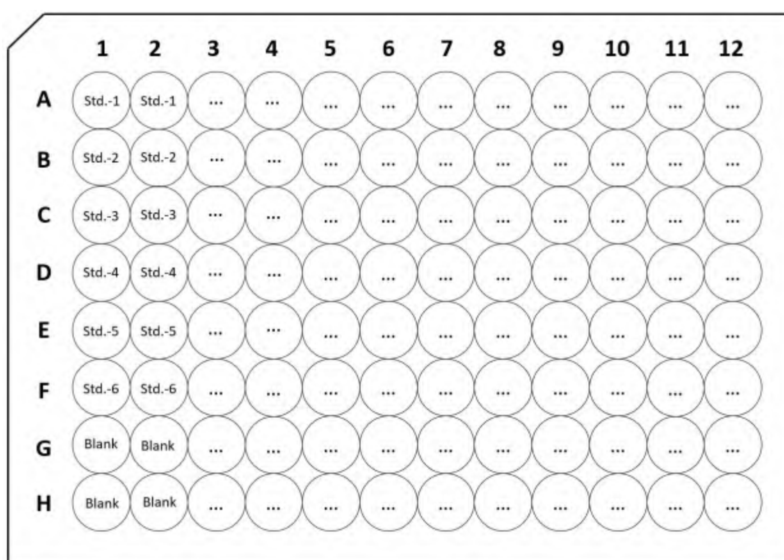
	Range (%)	83.0-99.0	82.1-102.7
1:16	Average Recovery (%)	88.7	94.9
	Range (%)	88.5-90.0	87.0-102.4

***Note**: The example data is for reference only.*

PLATE LAYOUT



***Note:** Blank is a Blank Dilution Buffer hole.*

TROUBLESHOOTING GUIDE

Problem	Cause	Solution
Poor standard curve	* Inaccurate pipetting	* Check pipettes
Large CV	* Inaccurate pipetting * Air bubbles in wells	* Check pipettes * Remove bubbles in wells
High background	* Plate is insufficiently washed * Contaminated wash buffer	* Review the manual for proper wash. * Make fresh wash buffer
Very low readings across the plate	* Incorrect wavelengths * Insufficient development time	* Check filters/reader * Increase development time

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Samples are reading too high, but standard curve looks fine	* Samples contain cytokine levels above assay range	* Dilute samples and run again
Drift	* Interrupted assay set-up * Reagents not at room temperature	* Assay set-up should be continuous - have all standards and samples prepared appropriately before commencement of theassay * Ensure that all reagents are at room temperature before pipetting into the wells unless otherwise instructed in the antibody inserts

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